Bi-Layered Living Cellular Constructs Mirrored Normal Skin and Accelerated Healing in a Porcine In Vivo Diabetic Delayed Wound Healing Model

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Introduction

Dysregulation and impairment in the process of normal wound healing may lead to chronic wounds. A number of skin substitutes are available to cover, support, and treat chronic wounds; however, the mechanism of these products remains largely unknown. In this study, we characterized key extracellular matrix (ECM) components, markers of activated keratinocytes, cellular proliferation, and key cytokines present in a bi-layered living cellular construct (BLCC^o) using immunofluorescence (IF) staining. Furthermore, a porcine *in vivo* diabetic delayed wound healing model was utilized to evaluate the impact of single or multiple applications of BLCC treatments using a porcine-derived BLCC.

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Methods

Structural assessments for BLCCs were made using hematoxylin and eosin (H&E) staining. To evaluate components relevant to normal skin, IF staining was completed for key ECM proteins (pro-collagen I, collagen I, collagen III, collagen IV, fibronectin, and laminin), markers of activated keratinocytes (K19), proliferation (Ki67), and key cytokines (VEGF, TGF- β , HGF, IGF-1). To assess the impact of BLCCs on wound healing, a porcine-derived BLCC (pBLCC) was developed and utilized in a diabetic delayed wound healing model. After chemical induction of diabetes, full-thickness wounds were created and then either left untreated (controls) or treated with single or multiple treatments of porcine BLCC.

Extracellular Matrix Proteins in BLCC



Figure 1: Extracellular matrix proteins present in BLCCs. A) Pro-collagen I, B) Collagen I, C) Collagen III, D) Collagen IV, E) Fibronectin, and F) Laminin. Blue = cell nuclei, orange = IF target. 20x magnification. Scale bar = 50μ m for all images.



Figure 2: Markers of activated keratinocytes, proliferation, and key cytokines present in BLCCs. A) Keratin 19, B) Ki67, C) VEGF, D) TGF- β , E) HGF, and F) IGF-1. Blue = cell nuclei, orange = IF target. 20x magnification. Scale bar = 50µm for all images.





Figure 3: Porcine BLCC showed comparable morphology as human analogs using H&E staining. A) Human BLCC is composed with intact epidermis (Epi) and dermal equivalent (DE). B) Porcine BLCC is comparable to human with stratified epidermal layers clearly visible (expansion).



Figure 4: In vivo full-thickness model. A) Study design and timeline. B) 3D rendering of the full-thickness wounds extending 0.5cm to the subcutaneous fat layer.



Conclusions

Treatment with pBLCC Improves Wound Healing

• BLCCs contain complex components and extracellular matrix proteins that are consistent with normal skin and previous immunohistochemistry of BLCC.

• Single and multiple (4x) treatments with pBLCC resulted in significant improvements in wound healing in an *in vivo* full-thickness porcine wound model; however, 4x BLCC resulted in significant differences earlier and at more time points, suggesting a greater impact from multiple treatments.

• Using predictive methods, 4x pBLCC treated animals are predicted to achieve ≥50% wound closure quicker than sham. 1x pBLCC treated animals are predicted to achieve $\geq 60\%$ wound closure quicker than sham.

 Increased wound closure was primarily driven by increased rate of reepithelization in response to pBLCC treatment in vivo.

