

(1) Critical Care Medicine Department, National Institutes of Health Clinical Center, Bethesda, MD, USA (3) Postdoctoral Research Associate Training Program, National Institute of General Medical Sciences, Bethesda, MD, USA (4) Laboratory of Transplantation Immunotherapy, National Institute of Dental and Craniofacial Research, National Institutes of Health, Bethesda, MD, USA (6) Laboratory of Vascular Thrombosis and Inflammation, National Heart, Lung, and Blood Institute, NIH, Bethesda, MD, USA (7) Laboratory of Infectious Diseases, National Institute of Allergy and Infectious Diseases, National Institutes of Health (NIH), Bethesda, MD, USA (8) Advanced Lung Transplant Program, Inova Fairfax Hospital, Falls Church, VA, USA (9) Laboratory of Immunoregulation, National Institute of Allergy and Infectious Diseases, National Institutes of Health (NIH), Bethesda, MD, USA

Background

- In Coronavirus Disease 2019 (COVID-19) neutrophilia and a hyperactive neutrophil response has been associated with worse outcomes as activated neutrophils provoke endothelial injury, coagulation, and progression to ARDS^{1,2,3}.
- COVID-19 induces an altered and exuberant antibody response, with titers and subclasses of antibodies correlating with disease severity⁴.
- Neutrophils respond to IgG and IgA antibody complexes, leading to a variety of effector functions such as phagocytosis, degranulation, and NETosis^{5,6}
- Here we query associations between the endogenous SARS-CoV-2 antibody response and neutrophil activation in COVID-19

Methods

- Baseline plasma samples from 57 patients hospitalized on oxygen with COVID-19 treated in a phase II study evaluating the safety and efficacy of fostamatinib, a spleen tyrosine kinase inhibitor, were obtained. We subsequently performed:
- Quantitative measurements of SARS-CoV-2 specific antibodies using a luciferase-based immunoprecipitation system assay
- Quantitative measurements of neutrophil specific biomarkers using Luminex technology
- Neutrophil extracellular traps (NETs) as measured by myeloperoxidase-DNA (MPO-DNA) complexes by ELISA.
- Absolute neutrophil count (ANC) and immature granulocyte count (IGC) were measured from complete blood counts (CBC).

Acknowledgements

This work was supported by the Division of Intramural Research (DIR) of the National Heart, Lung and Blood Institute, National Institute of Allergy and Infectious Disease and the Clinical Center of the US National Institutes of Health

SARS-CoV-2 Antibody Levels Associate with Neutrophil Activation

Richard W. Childs⁴, Daniel S. Chertow^{1,9}, Jeffrey R. Strich¹



Seth Warner^{1,9}, Rui Miao², Marcos J. Ramos-Benitez^{1,3}, Xin Tian², Robert Reger⁴, Peter D. Burbelo⁵, Yogendra Kanthi⁶, Jeffrey I. Cohen⁷, Anthony F. Suffredini¹, Steven D. Nathan⁸,



Results

Severe COVID-19 was associated with higher levels of nucleocapsid-IgA as well as spike-IgG compared to moderate disease

Levels of IgG-spike and IgG-nucleocapsid both had significant correlations with Absolute Neutrophil Count

All four antibody titers showed strong correlations with Immature Granulocyte count, evidence of emergency granulopoiesis and altered neutrophil populations

S100A9, a component of calprotectin, lactoferrin and lipocalin-2, components of primary and secondary granules correlated strongly with antibody titers

Circulating NETs levels demonstrated a significant correlation with spike IgA levels, and its correlations with IgG-spike and IgA-nucleocapsid additionally approached significance

Conclusion

Higher anti-spike and anti-nucleocapsid IgG and IgA levels associated with more severe COVID-19 illness.

Endogenous SARS-CoV-2 specific antibody levels associate with markers of emergency granulopoiesis and neutrophil hyperactivation

Circulating NETs levels, which have been previously been associated with disease severity and thrombotic markers, correlated with antibody levels⁷

Targeting antibody mediated neutrophil activation may prove beneficial to treatment of SARS-CoV-2⁸

References

Bonaventura, A., et al., Endothelial dysfunction and immunothrombosis as key pathogenic mechanisms in COVID-19. Nat Rev Immunoo, 2021. 21(5): p. 319-329

Reusch, N., et al., Neutrophils in COVID-19. Front Immunol, 2021. 12: p. 652470.

Meizlish ML, et al. A neutrophil activation signature predicts critical illness and mortality in COVID-19. Blood Adv. 2021 Mar 9;5(5):1164-1177

Garcia-Beltran, WF., COVID-19 neutralizing antibodies predict disease severity and survival. Cell. 2021 184(2):476-488 Stacey HD, et al., IgA potentiates NETosis in response to viral infection. Proc Natl Acad Sci U S A. 2021 Jul 6;118(27):e2101497118.

Wang Y, et al., Expression, Role, and Regulation of Neutrophil Fcy Receptors. Front Immunol. 2019 Aug 27;10:1958. Zuo Y, et al., Neutrophil extracellular traps in COVID-19. JCI Insight. 2020 Jun 4;5(11):e138999.

Strich, JR et al., Fostamatinib for the treatment of hospitalized adults with COVID: A randomized clinical trial, Clin Infec Dis 2021;ciab732