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In Vitro Activities of Epetraborole, a Novel Bacterial Leucyl-tRNA Synthetase Inhibitor, Against Mycobacterium avium Complex Isolates M.S. DeStefano¹ C.M. Shoen¹, M.R.K. Alley², M.H. Cynamon¹

ABSTRACT

Background: Epetraborole (EBO) is a boron-containing, oral inhibitor of bacterial leucyltRNA synthetase, an essential enzyme in protein synthesis; EBO demonstrates potent activity against nontuberculous mycobacteria. We evaluated the effects of select culture conditions on MIC determinations of EBO against isolates of M. avium complex (MAC), as well as EBO MIC₉₀ results with Middlebrook 7H9 broth compared to those with cation-adjusted Mueller Hinton Broth (CAMHB) for 51 MAC isolates.

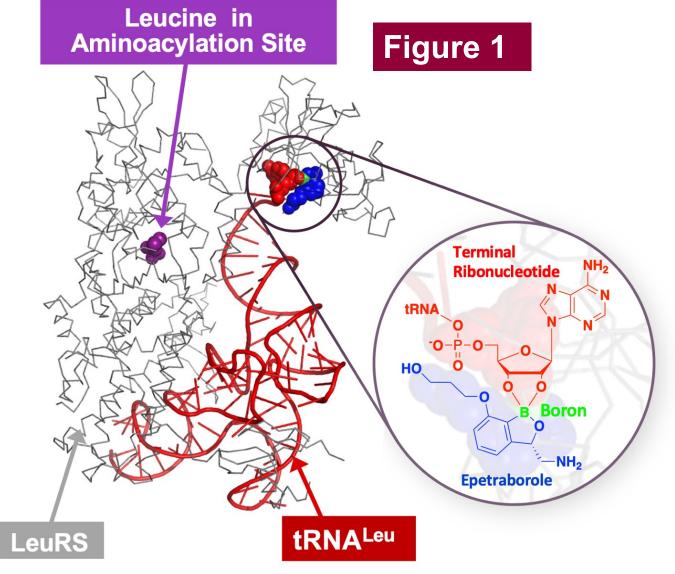
Methods: Six strains of MAC were used to test the *in vitro* activity of EBO in different conditions in a broth microdilution (BMD) assay. Activity was compared in Middlebrook 7H9 and CAMHB with 5% OADC from different manufacturers. The effects of glycerol, cations, oxyrase, varying pH levels, and increasing inoculum sizes were tested. Finally, EBO *in vitro* activity was tested for 51 MAC isolates in a BMD assay in both Middlebrook 7H9 and CAMHB with 5% OADC

Results: In general, manipulation of select culture conditions caused very little variation in EBO MIC values for the 6 MAC strains except for increasing the inoculum from ~10⁵ to 10⁷ CFU/mL, which caused an approximately 64x increase in the MIC. Since 1 MAC isolate out of 6 was affected by the addition of casitone, we tested 51 MAC isolates in both the minimal media Middlebrook 7H9 and the complex media CAMHB. EBO had a narrow MIC range in both broths, 0.25-8 mg/L for all isolates. The EBO modal MIC, MIC_{50} and MIC_{90} for the entire MAC panel of 51 isolates was 2 mg/L, 2 mg/L, and 8 mg/L for CAMHB and 1 mg/L, 1 mg/L, and 4 mg/L for Middlebrook 7H9, respectively **(Table 1)** Three clarithromycin-resistant isolates had EBO MIC values of 0.5 mg/L, 1 mg/L, and 2 mg/L suggesting that clarithromycin resistance does not affect EBO in vitro activity. In addition, amikacin resistance as determined using the Clinical Laboratory Standards Institute (CLSI) IV amikacin breakpoint (MIC ≥64 mg/L) had no noticeable effect on EBO MIC values

Conclusions: The MIC distribution for the 51 MAC isolates tested was similar in both media types, indicating that CAMHB can be used to test EBO MAC susceptibilities per CLSI guidelines. Clarithromycin- and amikacin-resistant isolates demonstrated no crossresistance with EBO.

INTRODUCTION

There are an estimated 200,000 patients with NTM lung disease in the United States with many remaining undiagnosed. The number of cases is increasing by an estimated 8% per year. Among the approximately 55,000 patients diagnosed with NTM lung disease in the United States approximately 44,000 patients have lung disease caused by MAC and approximately 35% of these patients have treatment-refractory MAC lung disease. Treatment of these infections is difficult due to the long courses of therapy that require a multiple drug regimen. This required course of treatment poses the challenges of patient non-adherence, expense, potential drug interactions, side-effects and/or adverse events, development of drug resistance, inferior outcomes and relapse or reinfection. EBO is a boron-containing, orally-available, small molecule inhibitor of bacterial leucyl-tRNA synthetase, an essential enzyme in protein synthesis¹ (**Figure 1**). EBO demonstrates potent activity against NTM². In this study, we evaluated the effects of select culture conditions on MIC determinations of EBO against isolates of MAC, as well as those with cation-adjusted Mueller Hinton Broth (CAMHB) for 51 MAC isolates.



Six strains of MAC were used to test the *in vitro* activity of EBO in different conditions using the broth microdilution (BMD) assay. Activity was compared in 7H9 and CAMHB with 5% OADC from different manufacturers. In addition, other conditions were tested including the addition of glycerol, using Chelex treated media plus cations³, oxygen depletion by the addition of Oxyrase, varying pH levels adding casitone (BD Acidicase[™] Peptone) and increasing the inoculum size. Finally, EBO *in vitro* activity was tested against 51 MAC isolates in a BMD assay in both 7H9 and CAMHB with 5% OADC

Table 1. Inoculum

Isol

M. avium ATCC 70

M. avium 2285R

M. intracellulare

M. intracellulare

M. intracellulare

MAC LPR ATCC 4

Table 2. The Effect

Strair

M. avium ATCC 70

M. avium 2285R

M. intracellulare

M. intracellulare D

M. intracellulare

MAC LPR ATCC 49

*Significant trailing was obs

REFERENCES

(1) Hernandez V et al. Discovery of a novel class of boron-based antibacterials with activity against Gram-negative bacteria. Antimicrob Agents Chemother. 2013;57:1394–1403. (2) Ganapathy US, Gengenbacher M, Dick T. Epetraborole Is Active against Mycobacterium abscessus. Antimicrob Agents Chemother. 2021 Sep 17;65(10):e0115621. (3) CLSI M100 ED30:2020. Performance Standards for Antimicrobial Susceptibility Testing, 30th edition.

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	suggesting that determined us			
Size Effect on I	MICs (mg/L) for	EBO in 7H9 + 5	% OADC	noticeable effe
te	~10 ⁵ /mL	~10 ⁶ /mL	~10 ⁷ /mL	Table 3. In
	inoculum	Inoculum	inoculum	Con
00898	1	L.) for EBO in 7H9 + 5% OADC nL ~10 ⁶ /mL ~10 ⁷ /mL inoculum inoculum inoculum 1 >64 1 >64 1 >64 4 >64 2 >64		
	0.25	1	>64	
ATCC 13950	0.5	1	>64	Epetraboro
DNA000111	1	4	>64	
1956	0.5	2	>64	
9601	0.5	2	>64	Clarithrom

		M				
	Drug	7H9	7H9 + casitone	САМНВ	Amikacin (AM	
	EBO	0.5	1	1		
0898	CLR	0.5	0.5	0.25		
	EBO	0.25	0.5	1		
	CLR	0.25	0.25	0.25		
ATCC 13950	EBO	0.5	0.5	0.5	• The MI	
	CLR	0.25	0.25	0.25	similar	
	EBO	2	*64 (4)	*64 (4)	 Based 	
DNA000111	CLR	2	2	1		
050	EBO	0.5	1	2	test EB	
956	CLR	0.5	1	0.5	• Clarith	
9601	EBO	0.5	1	1	demon	
	CLR	4	1	0.5		

RESULTS

In general, manipulation of select culture conditions caused very little variation in EBO MIC values for the 6 MAC strains except for increasing the inoculum from $\sim 10^5$ to 10^7 CFU/mL, which caused an approximately 64x increase in the MIC (Table 1). Since 1 MAC isolate out of 6 was affected by the addition of casitone (Table 2), we tested 51 MAC isolates in both the minimal media Middlebrook 7H9 and the complex media CAMHB. EBO had a narrow MIC range in both broths, 0.25-8 mg/L for all isolates. The EBO modal MIC, MIC₅₀ and MIC₉₀ for the entire MAC panel of 51 isolates was 2 mg/L, 2 mg/L, and 8 mg/L for CAMHB and 1 mg/L, 1 mg/L, and 4 mg/L for Middlebrook 7H9, respectively (Table 3). Three clarithromycin-resistant isolates had EBO MIC values of 0.5 mg/L, 1 mg/L, and 2 mg/L hat clarithromycin resistance does not affect EBO in vitro activity. In addition, amikacin resistance as sing the Clinical Laboratory Standards Institute (CLSI) IV amikacin breakpoint (MIC ≥64 mg/L) had no ect on EBO MIC values (Table 4).

ro Activity Against 51 Isolates of MAC					
ound	MIC Parameter (mg/L)	CAMHB + 5% OADC	7H9 + 5% OADC		
(EBO)	MIC Range	0.25-8	0.25-8		
	MIC Modal	2	1		
	MIC ₅₀	2	1		
	MIC ₉₀	8	4		
n (CLR)	MIC Range	0.25->64	0.25->64		
	MIC Modal	1	4		
	MIC ₅₀	1	2		
	MIC ₉₀	4	8		
IK)	MIC Range	8->64	8-32		
	MIC Modal	64	16		
	MIC ₅₀	16	16		
	MIC ₉₀	64	16		

CONCLUSIONS

The MIC distribution for the 51 MAC isolates tested was similar in 7H9 + 5% OADC and CAMHB + 5% OADC Based on the MIC results, CAMHB + 5% OADC can be used to test EBO MAC susceptibilities per CLSI recommendations Clarithromycin- and amikacin-resistant isolates demonstrated no cross-resistance with EBO

ACKNOWLEDGMENTS

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Table 4. MIC (mg/L) of EBO, CLR and AMK for 51 MAC Isolates						
Strain	CAMHB + 5% OADC		7H9 + 5% OADC			
	EBO	CLR	AMK	EBO	CLR	AMK
20-S-01 <i>M. chimaera</i>	4	1	16	1	1	16
20-S-02 <i>M. chimaera</i>	1	1	16	1	1	16
20-S-03 <i>M. chimaera</i>	1	1	16	0.5	1	16
20-S-04 <i>M. chimaera</i>	2	2	16	1	1	16
20-S-05 M. chimaera	8	2	64	4	4	16
20-S-06 <i>M. chimaera</i>	2	1	16	1	1	16
20-S-07 <i>M. chimaera</i>	2	1	16	1	1	16
20-S-08 <i>M. chimaera</i>	2	1	16	2	1	16
20-S-09 <i>M. chimaera</i>	0.5	0.5	8	0.5	0.5	16
20-S-10 <i>M. chimaera</i>	1	1	16	1 4	1	16
20-S-11 <i>M. intracellulare</i> 20-S-12 <i>M. intracellulare</i>	4 8	1	32 32	4 8	2	16 16
20-S-12 <i>M. Intracellulare</i> 20-S-13 <i>M. intracellulare</i>	o 2	>64	32 >64	° 2	2 >64	32
20-S-13 <i>M. Intracellulare</i>	Ζ Λ	-04	32	1	>04 0.5	3Z 16
20-S-14 <i>M. Intracellulare</i>	4 8	1	32	8	0.5	16
20-S-16 <i>M. avium hominissuis</i>	1	0.5	32	2	2	16
	1		-	2 1	4	_
20-S-17 <i>M. avium hominissuis</i>	•	0.5	64	•	-	16
20-S-18 <i>M. avium hominissuis</i>	0.25	0.25	16	0.25	4	16
20-S-19 <i>M. avium hominissuis</i>	0.5	0.5	64	1	4	16
20-S-20 <i>M. avium hominissuis</i>	2	1	16	1	2	16
20-S-21 <i>M. avium hominissuis</i>	0.5	0.5	64	1	4	16
20-S-22 M. avium hominissuis	8	4	16	8	8	16
20-S-23 M. avium hominissuis	0.5	0.5	64	1	4	16
20-S-24 <i>M. avium hominissuis</i>	4	2	32	4	4	16
20-S-36 <i>M. avium hominissuis</i>	4	2	>64	4	8	16
20-S-37 M. avium hominissuis	2	2	16	2	4	16
20-S-38 <i>M. avium hominissuis</i>	1	0.5	64	1	4	16
20-S-39 <i>M. avium hominissuis</i>	2	1	8	0.5	4	16
20-S-40 <i>M. avium hominissuis</i>	4	2	32	4	4	16
20-S-41 <i>M. avium hominissuis</i>	- 0.5	0.5	16	- 0.5	- - 1	16
20-S-42 <i>M. avium hominissuis</i>	2	4	16	2	4	16
	2	-	-	4	4	_
20-S-43 <i>M. avium hominissuis</i>	1	2	16	4	-	16
20-S-44 <i>M. avium hominissuis</i>	1	0.5	64	1	4	16
20-S-45 <i>M. avium hominissuis</i>	8	4	64	8	8	16
20-S-46 <i>M. intracellulare</i>	4	1	32	4	2	16
20-S-47 <i>M. intracellulare</i>	4	2	32	4	4	16
20-S-48 <i>M. intracellulare</i>	4	1	16	2	1	16
20-S-49 <i>M. intracellulare</i>	2	2	16 8	1	2	16 8
20-S-50 <i>M. intracellulare</i> 20-S-51 <i>M. intracellulare</i>	2	1	8 8	0.5	1	8 16
20-S-51 M. Intracellulare 20-S-52 M. intracellulare	4	1	8 16	0.5	1	16
20-S-52 M. Intracellulare	4 2	1	16	2	1	16
20-S-53 M. Intracellulare	2	1	8	2	4	16
20-S-54 M. Intracellulare	4	0.5	8 16	1	4 2	16
<i>A. avium</i> 2285R	4	2	8	0.25	0.5	16
M. intracellulare ATCC 13950	0.5	0.25	8 16	0.25	0.5	16
MAC 779	1	>64	32	0.5	>64	16
<i>I. intracellulare</i> 1956	0.5	0.25	8	0.5	0.25	8
MAC LPR ATCC 49601	0.5	0.25	32	0.5	4	16
MAC 623	0.5	>64	16	1	- >64	16
<i>I. intracellulare</i> 462	8	1	32	1	1	16
	0		VL			