

# Molecular diversity and resistance mechanisms of *Klebsiella pneumoniae* bloodstream infections in Peru

Fiorella Krapp<sup>1,2,3</sup>, Guillermo Salvatierra<sup>4</sup>, Noemí Hinojosa<sup>1</sup>, Coralith García<sup>1,2,5</sup>, Lizeth Astocondor<sup>1</sup>, Theresa Ochoa<sup>1,2</sup>, Jan Jacobs<sup>3,6</sup>, Omai Garner<sup>7</sup>, Víctor Nizet<sup>8</sup>, Pablo Tsukayama<sup>1,4</sup>

<sup>1</sup>Instituto de Medicina Tropical Alexander von Humboldt, Universidad Peruana Cayetano Heredia, Lima, Peru; <sup>2</sup>Facultad de Medicina, Universidad Peruana Cayetano Heredia, Lima, Peru; <sup>3</sup>Department of Microbiology, Immunology and Transplantation, KU Leuven, Belgium; <sup>4</sup>Laboratorio de Genómica Microbiana, Universidad Peruana Cayetano Heredia, Lima, Peru; <sup>5</sup>Hospital Nacional Cayetano Heredia, Lima, Peru; <sup>6</sup>Institute of Tropical Medicine Antwerp, Belgium; <sup>7</sup>Department of Pathology, University of California Los Angeles, USA; <sup>8</sup>Department of Pediatrics, University of California San Diego, USA.



## BACKGROUND

- Klebsiella pneumoniae*, is a leading pathogen for mortality associated with antimicrobial resistance (AMR), responsible for > 250,000 deaths in 2019.
- Genomic surveillance is gaining traction as an important tool for AMR surveillance, to identify high risk clones and emerging mechanisms of AMR.
- Important gaps currently exist in publicly available genomic data
  - Limited representativity of low- and middle-income countries
  - Limited epidemiological and clinical data linked to genomic data

## OBJECTIVES

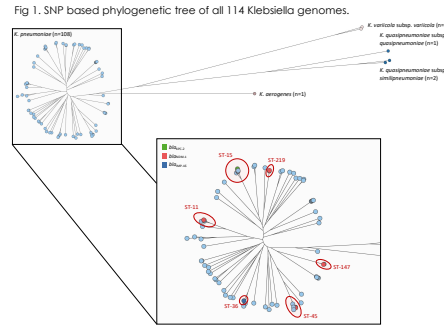
- Assess the genetic diversity among *K. pneumoniae* blood isolates recovered from different regions of Peru
- Identify the AMR and virulence genetic determinants of surveilled *K. pneumoniae* blood isolates
- Evaluate if epidemiological characteristics (age group, origin of infection, region) and clinical characteristics (severity and outcome) are associated with specific clonal groups (CGs), AMR genes and/or virulence genes.

## MATERIALS & METHODS

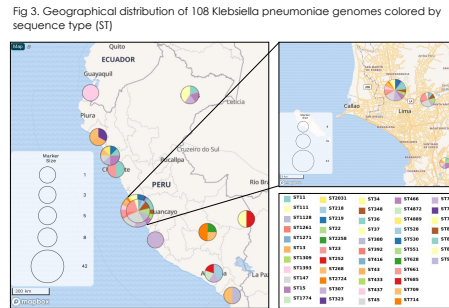
- Consecutive non-duplicate *K. pneumoniae* blood culture isolates collected during an AMR surveillance study (VIRAPERU), from Jul 2017 to Oct 2019, from 15 tertiary hospitals across Peru, were reactivated for whole genome sequencing.
- DNA extraction (GeneJET, Thermo Fisher Scientific), DNA library (Nextera XT, Illumina) and genome sequencing (MiSeq 500bp-V2, Illumina)
- De novo assembling (SPAdes v3.13.1), quality assessment and annotation (Nullarbor v2.0), and identification of species, sequence type (ST) group, K/O loci, and antimicrobial resistance genes (ARGs) and mutations (Kleborate v2.0.1)
- SNP trees were constructed using CSI Phylogeny 1.4 and visualized using Microreact.

## RESULTS

1 Six *Klebsiella* species were identified by WGS. The most prevalent was *K. pneumoniae sensu stricto* (n= 108, 95%).



From a total of 118 isolates, 114 were recovered and sequenced (4 did not grow).



2 Wide genetic diversity was observed among *K. pneumoniae* genomes along with a vast resistome.

- 54 ST groups
- 52 distinct K-loci
- 9 distinct O-loci
- 80 distinct ARGs
- 3 carbapenemase genes

Carriage of more than one ARG was present in 75.4% (86/114) *Klebsiella* genomes, with an average of 8.25 [95%CI: 0.52-7.22] ARGs per genome.

Isolates with a high number of virulence genes carried less ARGs.

Fig 2. SNP based phylogenetic tree of 108 *Klebsiella pneumoniae* genomes, along with their sequence type, K-loci and O-loci, virulence genes and ARGs and AMR mutations.



3 Carbapenem resistance was found in 13% of isolates. The most common carbapenemase gene was bla<sub>NDM-1</sub>.

Colistin resistance conferred by *mcr-1* was found in one isolate, with co-c carriage of ESBL gene bla<sub>CTX-M-15</sub>.

Table 1. Frequency of resistance to different antibiotics and the identified genetic determinants

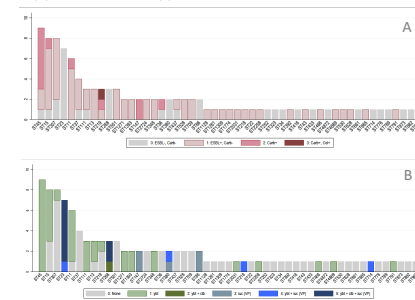
Resistance profile	Frequency of resistance	Related ARGs & alleles	Related AMR mutations	NP of isolates with no genetic determinant
Third-generation cephalosporins	77 (71.9%)	bla <sub>CTX-M-55</sub> (74.0%) bla <sub>CTX-M-26.1</sub> (2.1%) bla <sub>CTX-M-55</sub> (5.2%) bla <sub>CTX-M-14</sub> (2.8%) bla <sub>CTX-M-27</sub> (2.8%) bla <sub>SHV-12</sub> (2.8%) bla <sub>SHV-2</sub> (1.3%) bla <sub>SHV-45</sub> (1.3%)		1 (1.3%)
Carbapenems	14 (13.0%)	bla <sub>NDM-1</sub> (12.6%) bla <sub>OXA-2</sub> (1.3%) bla <sub>IMP-16</sub> (7.1%)		1 (7.1%)
Fluoroquinolones	70 (64.8%)	qnrB1 (34.2%) qnrB19 (28.0%) qnrS2 (15.7%) qnrS1 (5.7%) qnrB4 (1.4%) qnrB6 (1.4%) qnrS1 (1.4%) qnrS3 (1.4%)	Gyrase (48.6%) ParCII (44.3%) GyraseI (15.7%)	1 (1.4%)
Aminoglycosides	55 (50.9%)	aac(7)-Ib-x1-2 (37.3%) aac(3)-Iia-x1 (69.1%) strAa (54.5%) strBb (56.4%) aac(2) (45.8%) aac(1) (5.9%) aac(3)-I (2.7%) aph(4)-Ia (12.7%) aac(1)-Ib (10.9%) aac(5) (5.9%) aac(1)-Ia (8.1%) aac(1)-Ib (8.1%)		0 (0%)
Colistin	12 (11.3%)	mcr-1 (8.3%) pmrB (8.3%)		11 (91.7%)

ARGs, antimicrobial resistance genes; AMR, antimicrobial resistance  
\*Only the presence of *mcr-1* and *pmrB* or *pmrA* mutations was assessed

4 Eight high-risk ST groups were identified, based on their association with carbapenemase carriage (Figure 4A) or with high virulence (Figure 4B).

- Carbapenemase-carriage: ST45, ST219, ST147, ST15, ST11, ST36
- Highly-virulent: ST23, ST268

Fig 4. Distribution of ST groups based on Kleborate resistance score (A) and virulence score (B)



5 *K. pneumoniae* isolates recovered from bloodstream infections from the community had a higher virulence score and lower resistance score compared with those from hospital origin.

Hospital isolates had significantly higher number of ARGs than community isolates.

Table 2. Genomic features assessed by epidemiological characteristics and clinical outcome

Genomic feature	Community origin*		Hospital origin*		Adult		Discharge status	
	N=13	N=42	N=25	N=78	N=50	N=22	N=50	N=22
Genomic virulence score**	0 (0)	3 (7.1%)	0 (0)	1 (1.3%)	22 (44.0)	13 (59.0)	0 (0)	0 (0)
Genomic resistance score**	0 (0)	2 (15.4)	0 (0)	0 (0)	1 (2.0)	0 (0)	1 (2.0)	0 (0)
NP of antibiotic classes with genotypic resistance	0 (0)	1 (7.7%)	0 (0)	0 (0)	1 (2.0)	0 (0)	1 (2.0)	0 (0)
NP of antimicrobial resistance	0 (0)	10 (6.3%)	0 (0)	11 (14.1%)	10 (20.0)	11 (50.0)	10 (20.0)	11 (50.0)

## CONCLUSIONS

- Bloodstream infections in Peru are caused by a wide diversity of *K. pneumoniae* strains, carrying multiple AMR genes. Carbapenem resistance is principally a result of bla<sub>NDM-1</sub> carriage, found across 6 specific ST groups.
- Genomic surveillance of *K. pneumoniae* can be conducted in Peru following published genomic surveillance frameworks and using publicly available genomic tools.
- This study constitutes a benchmark for genomic surveillance of *K. pneumoniae* in Peru and a potential roadmap for other low-resource settings.

## ACKNOWLEDGEMENTS

This study was funded by the Fogarty International Center of the National Institutes of Health under Award Number D43TW009343 and the University of California Global Health Institute; the Young Investigator Award from the Institut Merieux; and the Belgian Directorate of Development Cooperation (DGD) through the Framework Agreement between the Belgian DGD and the Institute of Tropical Medicine, Belgium.

## CONTACT

Fiorella Krapp, MD, MSc  
Instituto de Medicina Tropical Alexander von Humboldt, Universidad Peruana Cayetano Heredia  
[fiorella.krapp@upch.pe](mailto:fiorella.krapp@upch.pe)  
[@fiorellakr](https://orcid.org/0000-0001-9148-1000)

## REFERENCES

- Antimicrobial Resistance Collaborators. 2022. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. *Lancet*. 399:629-655.
- GLASS whole-genome sequencing for surveillance of antimicrobial resistance. Geneva: World Health Organization; 2020
- Lam, M.M.C., Wick, R.R., Watts, S.C. et al. A genomic surveillance framework and genotyping tool for *Klebsiella pneumoniae* and its related species complex. *Nat Commun* 12: 4188 (2021).
- Wyrnes, K.L., Lam, M.M.C., & Holt, K.E. Population genomics of *Klebsiella pneumoniae*. *Nat Rev Microbiol* 18, 344-359 (2020).
- Wyrnes KL, Wick RR, Gorrie C, Jenney A, Follador R, Thomson NR, Holt KE. Identification of *Klebsiella* capsule synthesis loci from whole genome data. *Microb Genom*. 2016 Dec 12;12(12):e000102.