

Characterization of circulating clinical multi-drug resistant and methicillin resistant Staphylococcus aureus isolates in Peru

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INTRODUCTION

Staphylococcus aureus is one of the major threats to hospitalized patients given its ability to rapidly acquire resistance to multiple antibiotics. Indeed, S. aureus is in the High Priority list for research and development of new antibiotics declared by the World Health Organization [1].

Hospitalized individuals with longer lengths of admission have an increase of colonization by multidrug resistant and/or methicillin resistant *S. aureus* (MDR-SA and MRSA, respectively), which limits therapeutic options [2, 3].

RESULTS

We obtained 3103 bacterial pathogens from 8 different hospitals in Lima, of those 232 were identified as *S. aureus* from individuals between 2015 and 2018. We then selected 80/232 (35%) isolates that were resistant to ≥ 2 antibiotic classes and/or MRSA from for further characterization, of which 42.5% were obtained from the lower respiratory tract (Table 1).

RESULTS (Continued)



Dissemination of *S. aureus* clones in health-care settings is of high concern where over-the-counter use of antibiotics is widespread, and therefore antimicrobial susceptibility surveillance is necessary to identify trends in resistance. In addition, molecular surveillance is also of critical importance due to its inherent capability to provide an accurate description of the distribution of clinically important clonal *S. aureus* populations.

In Latin America, sequence type (ST) 5 clones have been associated with previous outbreaks [4], and it is currently one of the predominant clones present [5, 6].

Here we describe a four-year surveillance study of MDR-SA and MRSA in hospitals in Lima with further characterization by whole genome sequencing (WGS).

MATERIAL & METHODS

Bacterial isolates of nosocomial origin were prospectively collected between 2015 and 2018 from hospitalized patients from 8 hospitals in Lima, Peru. Sample's origin/type were categorized as: lower respiratory tract, upper respiratory tract, wound, blood, urinary tract, abdominal cavity, and rectal swabs. Bacterial isolates were transported from the hospital site to the NAMRU-6 laboratory and underwent further characterization, as follows:

Table 1. MDR-SA frequencies according to nosocomial origin.

Origin/type	Isolates	Frequency
Lower respiratory tract	34	42.5%
Upper respiratory tract	13	16.25%
Wounds	4	5%
Blood	17	21.25%
Urinary tract	2	2.5%
Abdominal cavity	5	6.25%
Rectal swab	5	6.25%
Total	80	100%

S. aureus isolates were resistant against nine different antimicrobial classes (Fig. 1). Of note, 79% (63/80) of MRSA isolates were also resistant to macrolides, quinolones, and lincomycins. Additionally, 81% (64/80) of the isolates had inducible resistance to clindamycin. Notably, 5% (4/77) of the MRSA isolates carried the Panton Valentine leucocidin gene *lukS* and the *mecA* gene, as revealed by multiplex PCR.

Figure 2. Frequencies of resistance associated genes to different antibiotic classes.







In silico MLST Sequence Types

Figure 3. MRSA and non-MRSA distribution in ST clusters.

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